

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
INSTITUTO DE CIÊNCIAS BÁSICAS DA SAÚDE
PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS BIOLÓGICAS:
FARMACOLOGIA E TERAPÊUTICA

Laís Pancotto

EFEITOS ANSIOLÍTICOS DE ACETIL-L-CARNITINA EM PEIXES-ZEBRA

Porto Alegre

2019

Laís Pancotto

EFEITOS ANSIOLÍTICOS DE ACETIL-L-CARNITINA EM PEIXES-ZEBRA

Dissertação apresentada ao Programa de Pós-Graduação em Ciências Biológicas: Farmacologia e Terapêutica do Instituto de Ciências Básicas da Saúde da Universidade Federal do Rio Grande do Sul como requisito parcial para a obtenção do título de mestrado em Farmacologia e Terapêutica.

Orientador: Prof. Dr. Angelo Piato

Porto Alegre

2019

CIP - Catalogação na Publicação

Pancotto, Laís
Efeitos ansiolíticos de acetil-L-carnitina em
peixes-zebra / Laís Pancotto. -- 2019.
45 f.
Orientador: Angelo Piato.

Dissertação (Mestrado) -- Universidade Federal do
Rio Grande do Sul, Instituto de Ciências Básicas da
Saúde, Programa de Pós-Graduação em Ciências
Biológicas: Farmacologia e Terapêutica, Porto Alegre,
BR-RS, 2019.

1. Acetil-L-carnitina. 2. Ansiedade. 3. Estresse
oxidativo. 4. Peixe-zebra. I. Piato, Angelo, orient.
II. Título.

Elaborada pelo Sistema de Geração Automática de Ficha Catalográfica da UFRGS com os
dados fornecidos pelo(a) autor(a).

AGRADECIMENTOS

Chegando ao final desta caminhada, primeiramente quero agradecer a Deus, que me guia em todos os momentos da minha vida. Gratidão aos professores e funcionários do PPG Farmacologia e Terapêutica pela acolhida, ensinamentos e amparo.

Ao professor Dr. Angelo Piato, meu sincero agradecimento por sua orientação, princípios, apoio e paciência ao longo do mestrado.

Agradecimento ao nosso grupo de pesquisa, em especial colegas Ricieri e Matheus que abriram mão de suas tarefas diárias para me auxiliar no laboratório.

À minha família (mãe, pai, irmão e avó), minha eterna gratidão pelo apoio, incentivo desde minha infância até hoje. Obrigada meu pai por sacrificar seu tempo para me ajudar em meus compromissos. Obrigada minha mãe pelo carinho e cuidado com minha bebê no tempo em que não me fiz presente.

Agradeço ao meu amor Douglas por entender meus objetivos, ser prestativo e fundamental em minha vida.

Gratidão a minha chefe e amiga Marinês Campestrini por tolerar minha ausência no trabalho e me incentivar na busca pelo conhecimento. Aos meus colegas de trabalho que me substituíam na farmácia básica quando não estava, ao querido companheiro de viagem e amigo Mauri, aos pacientes que entenderam minha ausência no meu trabalho.

As minhas colegas Véra e Ticiane pelo companheirismo, conselhos e amizade. Agradeço também Cláudia e Carla da Belo Farma Farmácia que aceitaram minhas divergências de horários ao longo do tempo.

E meu eterno e mais sincero agradecimento vai para minha filha Sarah. Meu presente de Deus, minha razão de viver, minha força para trabalhar além do horário, viajar por horas até chegar em Porto Alegre, superar todos os obstáculos, incertezas e dificuldades.

Filha, você me faz ser melhor a cada dia, me faz ter mil e uma utilidades, faz meu tempo dobrar, meus medos sumirem e minha bondade florescer. Se cheguei até o final foi por ti, minha amada filha.

Que os ensinamentos que recebi ao longo do mestrado me tornem uma pessoa melhor, humilde e humana. Que eu busque amenizar a dor do próximo não somente com medicamentos, mas também com cuidado, carinho e amor.

RESUMO

A acetil-L-carnitina (ALC) é a forma acetilada do aminoácido L-carnitina capaz de modular diversos alvos relacionados a fisiopatologia dos transtornos mentais. A ALC atravessa a barreira hematoencefálica de uma maneira mais eficaz do que a L-carnitina. Estudos demonstraram que a ALC possui atividade antioxidante, neuroprotetora e reguladora da neuroplasticidade, aumentando a expressão de BDNF e de receptores metabotrópicos de glutamato. Dessa forma, considerando o mecanismo multialvo da ALC, o objetivo deste estudo foi investigar os efeitos desse composto sobre parâmetros comportamentais e bioquímicos em peixes-zebra. Diferentes grupos de animais foram tratados com ALC (0,1, 1 e 10 mg/L) e posteriormente submetidos aos testes de tanque-novo, claro-escuro e estresse por perseguição. Animais controle não foram submetidos a nenhum tratamento, mas foram submetidos as mesmas condições experimentais. Posteriormente, os animais foram eutanasiados e os encéfalos coletados para a avaliação de parâmetros relacionados ao estresse oxidativo. ALC apresentou efeito ansiolítico no teste de tanque novo e de claro/escuro, além de prevenir o comportamento ansiogênico induzido por um estressor agudo (estresse por perseguição). Além disso, a ALC foi capaz de prevenir a lipoperoxidação induzida pelo estresse agudo no encéfalo dos peixes-zebra. Os dados apresentados aqui justificam a continuidade das investigações dos potenciais efeitos da ALC no tratamento de transtornos psiquiátricos relacionados ao estresse.

Palavras-chave: Acetil-L-carnitina. Ansiedade. Estresse oxidativo. Peixe-zebra.

ABSTRACT

Acetyl-L-carnitine (ALC) is the acetylated form of the amino acid L-carnitine capable of modulating various targets related to the pathophysiology of mental disorders. ALC crosses the blood-brain barrier more effectively than L-carnitine. Studies have shown that ALC has antioxidant, neuroprotective and neuroplasticity activity, increasing the BDNF and metabotropic glutamate receptors expression. Thus, considering the multi-target mechanism of ALC, the objective of this study was to investigate the effects of this compound on behavioral and biochemical parameters in adult zebrafish. Different groups of animals were treated with ALC (0.1, 1 and 10 mg/L) and subsequently submitted to the novel tank, light/dark and chasing stress tests. Control animals were not subjected to any treatment but were submitted to the same experimental conditions. After the behavioral tests, the animals were euthanized, and the brains were collected for the evaluation of parameters related to oxidative stress. ALC showed anxiolytic effect in the novel tank and light/dark tests, as well as was able to prevent anxiety-induced by chasing stress. In addition, ALC was able to prevent lipid peroxidation induced by acute stress in the zebrafish brain. The data presented here justify the continuation of investigations of the potential effects of ALC in the treatment of stress-related psychiatric disorders.

Keywords: Acetyl-L-carnitine. Anxiety. Oxidative stress. Zebrafish.

SUMÁRIO

1 INTRODUÇÃO	9
1.1 Revisão de literatura	9
1.2 Acetil-L-carnitina (ALC)	12
2 OBJETIVO	16
2.1 Objetivo geral	16
2.2 Objetivos específicos	16
3 ARTIGO CIENTÍFICO	17
4 DISCUSSÃO E CONCLUSÃO	36
REFERÊNCIAS	38

LISTA DE ABREVIATURAS, SÍMBOLOS E SIGLAS

5-HT Serotonina

6-OHDA 6-hidroxidopamina

AKT Proteína Cinase

ALC Acetil-L-carnitina

ATP Adenosina Trifosfato

BAX Regulador Apoptótico

BCL-2 Proteínas Antiapoptóticas

CAT Catalase

COMT Catecol-O-Metil Transferase

DNA Ácido Desoxirribonucleico

DSM-V Manual Diagnóstico e Estatístico de Transtornos Mentais 5ª Edição

EROs Espécies Reativas de Oxigênio

EUA Estados Unidos da América

GABA Ácido Gama-Aminobutírico

GSH Glutationa Reduzida

GSK 3b Glicogênio Sintase Cinase 3 beta

ISRS Inibidores Seletivos da Receptação de Serotonina

ISRSN Inibidores Seletivos da Receptação de Serotonina e da Noradrenalina

MAO Monoaminaoxidase

NAC N-acetilcisteína

NMDA N-metil D-Aspartato

NPSH Grupos Tióis não-proteicos

SOD Superóxido Dismutase

TBARS Substâncias Reativas ao Ácido Tiobarbitúrico

UCS Estresse Crônico Imprevisível

1 INTRODUÇÃO

A acetil-L-carnitina (ALC) é a forma acetilada do aminoácido L-carnitina capaz de modular diversos alvos relacionados a fisiopatologia dos transtornos mentais. Recentemente, estudos pré-clínicos e clínicos demonstraram os efeitos da ALC em parâmetros relevantes para ansiedade, esquizofrenia e transtornos do humor. Dessa forma, considerando o mecanismo multialvo da ALC, o objetivo deste estudo foi investigar os efeitos desse composto sobre parâmetros comportamentais e bioquímicos em peixes-zebra, agregando informações sobre os resultados obtidos em diferentes modelos de ansiedade em peixes-zebra, bem como sobre o potencial mecanismo de neuromodulação da ALC.

1.1 Revisão de literatura

O estresse pode ser definido como uma reação complexa desencadeada pelo organismo frente a estressores de origens variadas. Tal reação é extremamente importante para que o organismo responda adequadamente as demandas do dia a dia (Galvão-Coelho et al., 2015; McEwen and Morrison, 2013). Entretanto, quando essas demandas ultrapassam a capacidade adaptativa do organismo, isso pode predispor o indivíduo a doenças como hipertensão, diabetes, disfunções sexuais e cognitivas, depressão e ansiedade (Portugal et al., 2016).

De acordo com o Manual Diagnóstico e Estatístico de Transtornos Mentais (DSM-V), os transtornos de ansiedade incluem transtornos que compartilham características de medo e angústia excessiva e perturbações comportamentais. Os transtornos de ansiedade incluem transtorno de ansiedade generalizada, transtorno

de ansiedade social, fobias específicas e transtorno de ansiedade de separação. Enquanto o medo é a resposta emocional à ameaça iminente real ou percebida, a ansiedade é a antecipação de ameaça futura (Craske et al., 2009).

As bases neurobiológicas dos transtornos de ansiedade não estão completamente esclarecidas, mas diversos sistemas de neurotransmissores estão envolvidos (Griebel and Holmes, 2013; Holmes et al., 2003). Recentemente, pesquisas têm associado transtornos psiquiátricos com o estresse oxidativo, o qual pode ser definido como o desequilíbrio entre a formação e remoção de agentes oxidantes no organismo, decorrente da geração excessiva de espécies reativas de oxigênio (EROs) e/ou da diminuição de antioxidantes endógenos (Maes et al., 2011; Ng et al., 2008). Resultados recentes do nosso grupo mostraram que N-acetilcisteína, uma molécula que possui atividade antioxidante, apresenta atividade ansiolítica e anti-estresse em peixes-zebra (Marcon et al., 2016) e camundongos (Mocelin et al., 2015). Dessa forma, abordagens que envolvam diminuição e/ou prevenção de estresse oxidativo poderão contribuir com o tratamento de psicopatologias como ansiedade e depressão.

Os primeiros fármacos utilizados para o tratamento dos transtornos de ansiedade foram os benzodiazepínicos. Esses compostos ligam-se a um sítio específico no receptor de GABA_A, potencializando o efeito inibitório do neurotransmissor. Após a ligação do GABA ao seu receptor, ocorre abertura do canal de cloreto e influxo desse íon para o meio intracelular, inibindo diversos eventos pós-sinápticos de transdução de sinal. Os efeitos ansiolíticos são mediados pelos receptores GABA_A que contêm a subunidade α_2 , enquanto a sedação ocorre por meio da subunidade α_1 (Griebel and Holmes, 2013). Clinicamente os benzodiazepínicos apresentam a vantagem de ter um início de ação rápido, causando redução da

ansiedade, da agressividade ou até sedação, entretanto, podem apresentar efeitos colaterais. Um dos efeitos colaterais mais pronunciados desta classe de fármaco é a amnésia anterógrada. Tais efeitos têm sido demonstrados em animais e humanos (Andreatini et al., 2001; Levitan et al., 2013). Os benzodiazepínicos também possuem um potencial de causar dependência. A exposição crônica provoca modificação na neurotransmissão gabaérgica que contribui para o desenvolvimento de tolerância, dependência e síndrome de abstinência (Nordon et al., 2009; Schalleberger et al., 2016).

Atualmente, o tratamento farmacológico considerado de primeira linha para transtornos de ansiedade são os antidepressivos inibidores seletivos da receptação de serotonina (ISRS) e inibidores seletivos da receptação de serotonina e da noradrenalina (ISRSN) (Griebel and Holmes, 2013). Os ISRS atuam no neurônio pré-sináptico inibindo especificamente a recaptação de serotonina. O aumento da disponibilidade sináptica de serotonina estimula a função de receptores (5-HT) pós-sinápticos, e acredita-se que a estimulação desses receptores contribui para os efeitos adversos característicos dessa classe de fármacos, incluindo efeitos gastrointestinais (náuseas, vômitos) e sexuais (demora ou comprometimento do orgasmo). A fluoxetina, a paroxetina, a sertralina e o escitalopram são exemplos de fármacos dessa classe. Dos fármacos ISRSN, a venlafaxina foi mais amplamente estudada, mas a duloxetina também já apresenta eficácia comprovada. No entanto, os ensaios clínicos com ISRS e ISRSN mostram 60 a 70% de resposta terapêutica. Além disso, em pacientes que fazem uso prolongado de ISRS ou ISRSN, a disfunção sexual e o embotamento afetivo se apresentam como limitações importantes (Amaral, 2014; Braga et al., 2010; Castillo et al., 2000).

Apesar da ampla variedade de tratamentos disponíveis para os transtornos de ansiedade, os efeitos adversos relacionados ainda causam problemas relevantes e contribuem para a baixa adesão ao tratamento. Relevante para este projeto estima-se que em torno de 50% dos pacientes sejam refratários a esses tratamentos, justificando o grande interesse no desenvolvimento de novos fármacos mais efetivos. É provável que esse objetivo só seja alcançável com o desenvolvimento de fármacos com mecanismo de ação inovador.

1.2 Acetil-L-carnitina (ALC)

O uso de suplementos nutricionais tem crescido ao longo das últimas décadas. Entre as substâncias que têm recebido grande atenção, destaca-se a ALC que é popularmente utilizada como coadjuvante na redução de gordura corporal e como termogênico (Brass, 2000; Silva et al., 2012).

A carnitina, um nutriente essencial da dieta, atua como carreador de ácidos graxos para o interior da mitocôndria onde ocorre a β -oxidação. Altas concentrações de carnitina podem ser encontradas em tecidos e células nas formas livre ou como acilcarnitinas, como a acetil-L-carnitina (Steiber et al., 2004). ALC possui inúmeras funções biológicas como auxiliar a captação de acetil-CoA na mitocôndria durante a oxidação dos ácidos graxos, estimular a produção de acetilcolina e a síntese de proteínas e de fosfolípidios, prevenindo a morte neuronal (Di Cesare Mannelli et al., 2010; Jones et al., 2010).

Além disso, a ALC possui atividade neuroprotetora e antioxidante (Chapela et al., 2009; Jones et al., 2010). A administração de ALC aumenta os níveis celulares de GSH em astrócitos de rato, facilita o transporte de GSH através da barreira

hematoencefálica (Kido et al., 2001) e eleva os níveis de GSH na via nigroestriatal (Fariello et al., 1988). Esse aumento dos níveis de GSH poderia proteger tanto sinaptossomas quanto mitocôndrias do estresse oxidativo mediado por espécies reativas de oxigênio (Drake et al., 2002). A administração da ALC diminuiu a lipoperoxidação em encéfalos de ratos idosos, além de prevenir a formação de radicais livres e a oxidação de proteínas em córtex frontal dos cães submetidos a isquemia cerebral e reperfusão (Calvani and Arrigoni-Martelli, 1999). A ingestão crônica oral de 500 mg/kg de ALC por roedores por 25 dias alterou o metabolismo energético cerebral e provocou aumento significativo nas concentrações de serotonina no córtex e noradrenalina no hipocampo de ratos (Smeland et al., 2012). A suplementação crônica de ALC causou diminuição do metabolismo de glicose em lactato, indicando aumento nos níveis de energia em algumas regiões do encefálicas (Smeland et al., 2012).

Estudos mostraram que a ALC aumentou a expressão do receptor metabotrópicos mGlu2 no córtex pré-frontal e hipocampo por meio de mecanismos epigenéticos mediados por acetilação das histonas e do fator de transcrição NFkb, possuindo potencial para o tratamento de transtornos do humor e de ansiedade, com um bom perfil de tolerabilidade. Além disso, ALC aumentou a neurogênese em camundongos submetidos ao protocolo de estresse crônico imprevisível (Cuccurazzu et al., 2013; Nasca et al., 2013). ALC também mostrou eficácia terapêutica sobre sintomas depressivos em pacientes com fibromialgia (Leombruni et al., 2015) e em pacientes idosos (Pettegrew et al., 2002).

Em outro estudo realizado em ratos, a administração crônica de ALC nas doses de 50 e 75 mg/kg, mas não 10 e 100 mg/kg, mostrou efeito sobre a ansiedade no teste de labirinto em cruz elevado (Levine et al., 2005). Recentemente, Lau e

colaboradores (2016) demonstraram que o tratamento por via oral durante três dias com ALC preveniu o aumento de ansiedade induzido por um protocolo de estresse por contenção no teste de claro escuro em ratos (Lau et al., 2016).

O tratamento com L-carnitina associado a uma dieta hipoproteica é capaz de conferir proteção contra o dano oxidativo a biomoléculas (lipídios, proteínas, enzimas e DNA) (Jones et al., 2010). Levando em consideração que o sistema nervoso central é extremamente suscetível ao estresse oxidativo devido ao grande conteúdo lipídico, intervenções capazes de prevenir e/ou reverter tais insultos poderiam ser utilizados no tratamento de transtornos psiquiátricos (Wang et al., 2014).

O antagonista do receptor NMDA cetamina reduziu os níveis de ATP em embriões peixes-zebra. Além disso, diminuiu a proteína mitocondrial e o potencial de membrana, causando deficiência de ATP mediada por disfunção mitocondrial. A maior parte do ATP gerado pelas mitocôndrias cardíacas é utilizada para sua contração e relaxamento. Embriões tratados com cetamina mostraram estrutura cardíaca anormal, comprometendo o desenvolvimento do coração. Essa atividade foi abolida com o co-tratamento com ALC, prevenindo os efeitos deletérios induzidos pela cetamina (Robinson et al., 2018). Um estudo também mostrou os efeitos da ALC sobre o comportamento e estresse oxidativo em peixes-zebra submetidos a um protocolo de estresse crônico imprevisível (UCS). O UCS induziu comportamento ansiogênico e estresse oxidativo enquanto o tratamento com ALC preveniu tais efeitos (Marcon et al., 2019).

Recentemente, uma equipe que incluiu pesquisadores de diversas instituições como a Universidade Rockefeller (EUA), a Universidade de Duke (EUA) e o Instituto Karolinska (Suécia), conduziu um experimento com roedores que concluiu que a ALC tinha um efeito antidepressivo de ação rápida nos animais e acredita-se que seu

mecanismo de ação impeça o disparo excessivo de neurônios excitatórios, mas essa função precisará ser mais explorada. A equipe também recrutou 71 pacientes com diagnóstico de depressão, homens e mulheres com idades entre 20 e 70 anos. Também recrutaram 45 pessoas saudáveis como grupo de controle. Quando comparados ao grupo de controle pareado por idade e sexo, os pacientes com depressão tinham níveis sanguíneos substancialmente mais baixos de ALC. Esses resultados sugerem que a ALC pode ser um potencial marcador biológico no transtorno depressivo maior (Nasca et al., 2018).

A ALC é uma molécula multialvo capaz de modular diversas vias relacionadas à fisiopatologia dos transtornos psiquiátricos relacionados ao estresse. Entretanto, ainda são poucos os estudos que mostram os efeitos desse composto em modelos animais de ansiedade. Dessa forma, a proposta desse trabalho foi avaliar os efeitos de ALC sobre parâmetros comportamentais e bioquímicos em peixes-zebra a fim de contribuir para a elucidação dos efeitos desse composto sobre o sistema nervoso central bem como estabelecer os possíveis mecanismos relacionados.

2 OBJETIVOS

2.1 Objetivo geral

Investigar os efeitos da exposição aguda à acetil-L-carnitina (ALC) sobre parâmetros comportamentais e bioquímicos em peixes-zebra.

2.2 Objetivos específicos

São objetivos específicos desta pesquisa:

- a) avaliar os efeitos da ALC nos testes de tanque novo e claro-escuro em peixes-zebra;
- b) investigar os efeitos da ALC sobre o comportamento e parâmetros de estresse oxidativo em peixes-zebra submetidos a um protocolo de estresse agudo (perseguição por rede).

3 ARTIGO CIENTÍFICO

Artigo publicado no periódico PeerJ. 2018 Jul 31;6:e5309.

doi: 10.7717/peerj.5309. eCollection 2018.

Anxiolytic and anti-stress effects of acute administration of acetyl-L-carnitine in zebrafish.

Lais Pancotto^{1#}, Ricieri Mocelin^{2#}, Matheus Marcon², Ana P. Herrmann¹, Angelo Piato^{1,2,3*}

¹Programa de Pós-Graduação em Farmacologia e Terapêutica, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil.

²Programa de Pós-Graduação em Neurociências, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil.

³Zebrafish Neuroscience Research Consortium (ZNRC), Slidell, LA, USA.

#Both authors contributed equally to this article.

Anxiolytic and anti-stress effects of acute administration of acetyl-L-carnitine in zebrafish

Lais Pancotto^{1,*}, Ricieri Mocelin^{2,*}, Matheus Marcon², Ana P. Herrmann¹ and Angelo Piato^{1,2,3}

¹ Programa de Pós-Graduação em Farmacologia e Terapêutica, Universidade Federal do Rio Grande do Sul, Porto Alegre, RS, Brazil

² Programa de Pós-Graduação em Neurociências, Universidade Federal do Rio Grande do Sul, Porto Alegre, RS, Brazil

³ Zebrafish Neuroscience Research Consortium (ZNRC), Los Angeles, United States of America

* These authors contributed equally to this work.

ABSTRACT

Studies have suggested that oxidative stress may contribute to the pathogenesis of mental disorders. In this context, molecules with antioxidant activity may be promising agents in the treatment of these deleterious conditions. Acetyl-L-carnitine (ALC) is a multi-target molecule that modulates the uptake of acetyl-CoA into the mitochondria during fatty acid oxidation, acetylcholine production, protein, and membrane phospholipid synthesis, capable of promoting neurogenesis in case of neuronal death. Moreover, neurochemical effects of ALC include modulation of brain energy and synaptic transmission of multiple neurotransmitters, including expression of type 2 metabotropic glutamate (mGlu2) receptors. The aim of this study was to investigate the effects of ALC in zebrafish by examining behavioral and biochemical parameters relevant to anxiety and mood disorders in zebrafish. ALC presented anxiolytic effects in both novel tank and light/dark tests and prevented the anxiety-like behavior induced by an acute stressor (net chasing). Furthermore, ALC was able to prevent the lipid peroxidation induced by acute stress in the zebrafish brain. The data presented here warrant further investigation of ALC as a potential agent in the treatment of neuropsychiatric disorders. Its good tolerability also subsidizes the additional studies necessary to assess its therapeutic potential in clinical settings.

Submitted 28 February 2018

Accepted 26 June 2018

Published 31 July 2018

Corresponding author

Angelo Piato, angelopiato@ufrgs.br

Academic editor

Pedro Silva

Additional Information and
Declarations can be found on
page 12

DOI 10.7717/peerj.5309

© Copyright
2018 Pancotto et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Subjects Neuroscience, Pharmacology

Keywords Acetyl-L-carnitine, Anxiety, Oxidative stress

INTRODUCTION

Acetyl-L-carnitine (ALC) facilitates the movement of acetyl-CoA into the mitochondria during the oxidation of fatty acids in mammals (*Chapela et al., 2009*). Moreover, this molecule is widely consumed as a dietary supplement for physical exercise (*Ribas, Vargas & Wajner, 2014; Nicassio et al., 2017*). Recently, preclinical and clinical studies have demonstrated the effects of ALC on parameters relevant to anxiety, schizophrenia, and mood disorders; with onset of action faster than antidepressant drug and exert

neuroprotective, neurotrophic, and analgesic effects (Levine et al., 2005; Wang et al., 2015; Traina, 2016; Singh et al., 2017; Nasca et al., 2017; Chiechio, Canonico & Grilli, 2017).

A growing body of evidence suggests that psychiatric disorders such as anxiety and depression are associated with oxidative damage (Ortiz et al., 2017; Niedzielska et al., 2016; Schiavone, Colaianna & Curtis, 2015; Cobb & Cole, 2015; Ng et al., 2008), since a decrease in antioxidant capacity can impair the organism's protection against reactive oxygen species and cause damage to fatty acids, proteins, and DNA (Maes et al., 2011). Superoxide and hydroxyl radical (free radicals) or hydrogen peroxide and their derivatives (non-radical molecules) called reactive oxygen species (ROS) are responsible for causing oxidative damage (Smaga et al., 2015). The antioxidant defense mechanism they are the non-enzymatic (i.g. glutathione) and enzymatic antioxidants (i.g. superoxide dismutase and catalase) which show a trend to decrease in neuropsychiatric diseases (Ozcan et al., 2004; Hassan et al., 2016). Preclinical and clinical research has evaluated antioxidant compounds (i.g. N-acetylcysteine, resveratrol and curcumin) in the treatment of psychiatric disorders, and it has been reported that these compounds are able to protect against oxidative stress-induced neuronal damage, preventing lipid peroxidation and behavioral changes (Mecocci & Polidori, 2012; Berk et al., 2014; Wang et al., 2014; Mocelin et al., 2015; Patel, 2016; Santos et al., 2017).

With simple, rapid and cheaper tests when compared with rodents, zebrafish have been used as a powerful complementary model for the study of a variety of neuropsychiatric diseases through behavioral and biochemical parameters (Stewart et al., 2015; Mocelin et al., 2015; Marcon et al., 2016; Marcon et al., 2018; Khan et al., 2017). There are several behavioral protocols extensively used and described for this species, such as the novel tank and light/dark tests. The novel tank diving test is based on an anti-predatory defense mechanism that induces fish to swim at the bottom of the tank, whereas the light/dark test evaluates anxiety based on the innate preference of adult zebrafish to dark over light areas (Levin, Bencan & Cerutti, 2007; Gebauer et al., 2011; Khan et al., 2017; Pittman & Piato, 2017).

In addition to its role in lipid metabolism, ALC also possesses free radical scavenging properties, and may thus protect the cells from oxidative damage by acting as an antioxidant (Gülçin, 2006; Sepand et al., 2016). Therefore, the aim of this study was to investigate the effects of ALC in zebrafish by examining behavioral and biochemical parameters relevant to anxiety and mood disorders in zebrafish.

MATERIALS AND METHODS

Animals

A total of 240 adult zebrafish (*Danio rerio*, F. Hamilton 1822) wild-type short fin strain (6-month-old, 3–4 cm long) 50:50 male/female ratio were purchased from Delphis aquariums (Porto Alegre, Brazil). The fish were kept for 15 days in a closed acclimation tank system of 16 L (40 × 20 × 24 cm) identical to the experimental tanks. Housing conditions consisted only of a tank with water, heater, filter and aeration system, and were maintained as previously described in Marcon et al. (2016). The tanks contained non-chlorinated, aerated

tap water (pH 7.0 ± 0.3 ; temperature 26 ± 1 °C; total ammonia at <0.01 mg/L; nitrite <0.01 mg/L; dissolved oxygen at 7.0 ± 0.4 mg/L; alkalinity at 22 mg/L CaCO₃ and total hardness at 5.8 mg/L), with a light/dark cycle of 14/10 h (lights on at 06:00 am). The fish were fed twice a day with a commercial flake fish food (Alcon BASIC®; Alcon, São Paulo, Brazil). On the experimental days, all the fish were only fed early in the morning before behavioral testing began. The order of testing was counterbalanced so that fasting time was randomized across experimental groups. All experiments were approved by the Ethics Committee of Universidade Federal do Rio Grande do Sul (#30992/2015).

Drug and experimental design

O-Acetyl-L-carnitine hydrochloride (ALC, CAS number 5080-50-2) was acquired from Sigma-Aldrich (St Louis, Missouri, USA). In all experimental protocols (novel tank, light/dark, and acute chasing stress tests), the animals were treated or not with ALC (0.1, 1.0 and 10.0 mg/L) in a beaker for 10 min. In the first protocol, immediately after the treatment, the animals were placed in the novel tank test (NTT) for 6 min. In the second protocol, after the treatments, the animals were placed in the light/dark test (LDT) for 5 min. Finally, in the third protocol, the animals were treated as previously and then chased with a net for 2 min. Then, the animals were placed in the NTT. The biochemical analyses were performed in animals submitted to this last protocol. A control group was submitted to the same experimental conditions (stressed or not) but without treatment. Different sets of animals were used in each experimental protocol. The experimental design is shown in Fig. 1 and was based on the previously published study by *Mocelin et al. (2015)*. All behavioral tests were performed between 09:00 am and 16:00 pm. The researchers who performed the behavioral tests and analyzed the data were unaware of the allocation of animals to the experimental groups. The concentrations were based on previous studies with another antioxidant compound (N-acetylcysteine) and pilot studies with a wider concentration range. The same concentrations were used in a chronic study with ALC (M Marcon, R Mocelin, A Araujo, A Herrmann & A Piato, 2018, unpublished data). We do not attempt to extrapolate the drug concentrations we used in a fish study to human dosage since there is not a straightforward calculation to be done. Since the half-life and other pharmacokinetic parameters of ALC in zebrafish are not known, it is difficult to precisely compare the concentration range that we observed here with the dose range for humans.

Novel tank test (NTT)

The novel tank test followed the protocol already described in *Mocelin et al. (2015)*. Briefly, the animals were separately moved to the apparatus (2.7-L tank, 24 × 8 × 20 cm, virtually divided into three equal horizontal zones and filled with standard tank water up to 15 cm) and video recorded for 6 min to be later analyzed by the ANY-maze™ software (Stoelting Co., Wood Dale, IL, USA). To evaluate exploratory behavior and locomotion we measured the parameters: total distance moved (m), number of transitions between zones, time spent in the upper and bottom zones of the tank, and number of transitions to the upper zone. Total distance and crossings were used as an indicator of overall locomotor activity. The

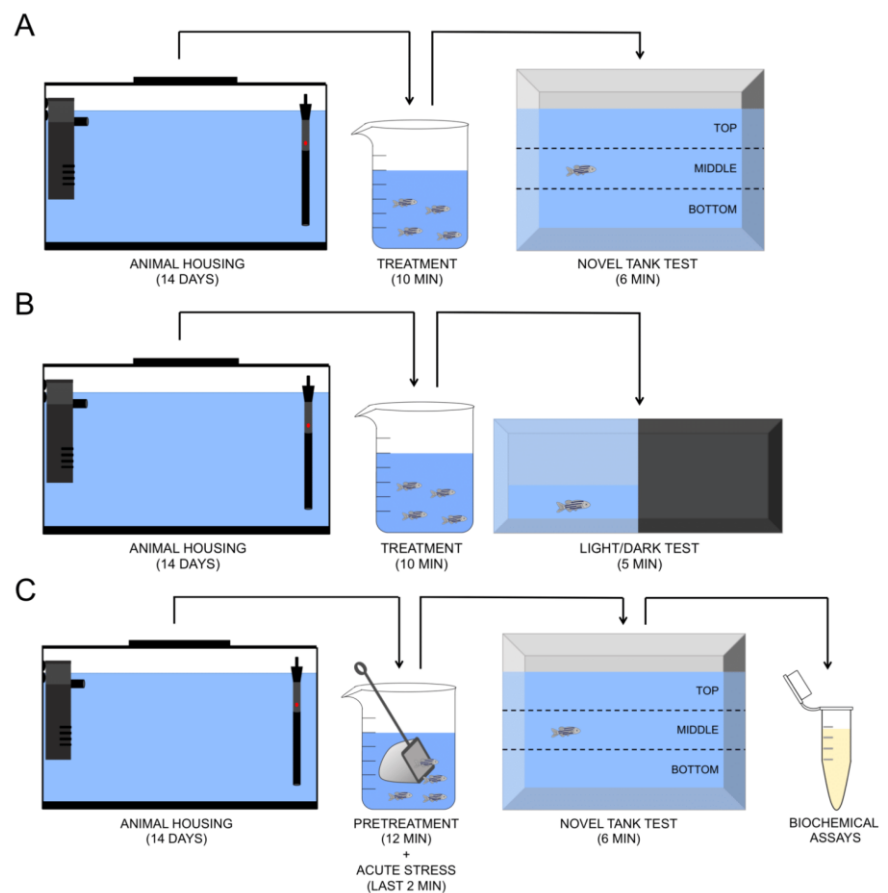


Figure 1 Schematic representation of the experimental protocol. Novel tank test (A), light/dark test (B), and acute chasing stress and biochemical assays (C).

Full-size [DOI: 10.7717/peerj.5309/fig-1](https://doi.org/10.7717/peerj.5309/fig-1)

upper zone of the tank corresponds in rats and mice protocols to the periphery region of the open-field test. Alterations in time spent and number of crossings to this zone are frequently used as a parameter of anxiety in zebrafish (Mocelin et al., 2015; Giacomini et al., 2016; Marcon et al., 2016; Mocelin et al., 2017; Marcon et al., 2018).

Light/dark test (LDT)

The light/dark test followed the protocol already reported by Gebauer et al. (2011). Specifically, the apparatus consisted of a glass tank (18 × 9 × 7 cm) divided by a raised glass into a dark and a white compartment of equal sizes, with the water level set at 3 cm and the partition raised 1 cm above the tank floor. One fish at a time was positioned in the white zone of the apparatus immediately after treatment. We recorded the number of

crossings and the time spent in the white compartment for 5 min. Zebrafish have a natural preference for dark environments and the white compartment is very anxiogenic for this species; anxiolytics increase the time spent in the white compartment (*Maximino et al., 2010; Mocelin et al., 2015*).

Acute chasing stress test (ACS)

The acute stress protocol was performed according to the previous study published by *Mocelin et al. (2015)*. Briefly, the animals were treated for 12 min and then chased for the last 2 min with a net before being moved to the novel tank, where they were recorded for 6 min. The behavioral parameters were quantified as described above for the NTT.

Tissue preparation

Samples were collected and prepared as previously reported by *Mocelin et al. (2018)*. Specifically, after the ACS fish were anesthetized by immersion in cold water and euthanized by decapitation. Each independent sample was then obtained by pooling four brains, which were homogenized on ice in 600 μ L phosphate buffered saline (PBS, pH 7.4; Sigma-Aldrich, St. Louis, MO, USA). The homogenate was centrifuged at 10,000 g for 10 min at 4 °C in a cooling centrifuge, and the supernatant was packed in microtubes for further assays.

Protein determination

Protein was determined by the Coomassie blue method described in detail by Bradford (1976). Specifically, we used bovine serum albumin (Sigma-Aldrich, St. Louis, MO, USA) as standard and the absorbance of samples was measured at 595 nm.

Lipid peroxidation (TBARS)

Lipid peroxidation was measured by the quantification of thiobarbituric acid reactive species (TBARS) production according to the method reported by (*Draper & Hadley, 1990*). More specifically, we followed the protocol described by *Mocelin et al. (2018)*, in which 50 μ L of the sample (80–100 μ g protein) was mixed with 75 μ L of trichloroacetic acid (TCA 10%; Sigma-Aldrich, St. Louis, MO, USA) and centrifuged at 6,000 rpm for 5 min at 4 °C in a cooling centrifuge. In the supernatants were added to 75 μ L thiobarbituric acid (TBA 0.67%; Sigma-Aldrich, St. Louis, MO, USA), then homogenized in a vortex for 5s and heated at 100 °C for 30 min. TBARS levels were measured by absorbance (532 nm) in a microplate reader, using malondialdehyde (MDA; Sigma-Aldrich, St. Louis, MO, USA) as a standard, and results were expressed as nmol MDA/mg protein.

Reduced thiol (SH) and Non-protein thiols levels (NPSH)

SH and NPSH levels were determined and measured at 412 nm in a microplate reader according to the method described by *Ellman (1959)*. More specifically, we followed the steps described by *Mocelin et al. (2018)*. Briefly, for SH the samples (60–80 μ g protein) were added to 10 mM 5,5-dithio-bis-2-nitrobenzoic acid (DTNB) dissolved in ethanol, developing yellow color after 1 h. The NPSH were similarly assessed, except that the sample was mixed with equal volumes of the 10% trichloroacetic acid (TCA) and centrifuged (6,000 rpm, 5 min). The supernatant was used for the biochemical assay. Results were expressed as μ mol SH/mg protein.

Superoxide dismutase (SOD) and catalase (CAT) activities

SOD and CAT activities were determined according to the method reported by *Misra & Fridovich (1972)* and *Aebi (1984)*, respectively. The protocol followed the more specific details described by *Dal Santo et al. (2014)*. Specifically, SOD activity was quantified in a microplate reader (480 nm) by testing the inhibition of radical superoxide reaction of the sample (20–30 μg protein) in the presence of adrenalin, monitoring adrenochrome formation in a medium containing a glycine-NaOH buffer (pH 10) and adrenalin (1 mM). CAT activity was assessed by measuring the decrease in H_2O_2 absorbance in a microplate reader (240 nm). The assay mixture consisted of sample (20–30 μg protein), phosphate buffered saline (pH 7.4), and 5 μL H_2O_2 (0.3 M). Results were expressed as units/mg protein.

Statistics

Normality and homogeneity of variance of the data were checked by D'Agostino-Pearson and Levene tests, respectively. Results were analyzed by one- or two-way ANOVA followed by Tukey's post hoc test. Two-way ANOVA was used to identify the main effects of stress and treatment, as well as their interactions. Data are expressed as a mean + standard error of the mean (S.E.M.). The level of significance was set at $p < 0.05$.

RESULTS

Behavioral parameters

Figure 2 shows the effects of ALC (0.1, 1.0 and 10.0 mg/L) on the novel tank test in zebrafish. ALC significantly increased the time spent in the top (0.1 and 1.0 mg/L, *Fig. 2D*) and decreased the time spent the bottom (0.1 mg/L, *Fig. 2E*) zone of the tank ($F_{3,77} = 8.0$, $p = 0.0001$ and $F_{3,77} = 5.6$, $p = 0.0016$, respectively). Locomotor parameters of groups treated with ALC (0.1, 1.0, and 10.0 mg/L) did not differ from control (*Figs. 2A* and *2B*).

In the light/dark test, ALC (0.1 and 10.0 mg/L) significantly increased the time spent in the lit side of the tank when compared to control ($F_{3,92} = 3.6$, $p = 0.0161$, *Fig. 3B*). The number of crossings between the light and dark compartments was not altered by any of the concentrations ($F_{3,92} = 0.9$, $p = 0.4284$, *Fig. 3A*).

Figure 4 shows the effects of ALC in the acute chasing stress (ACS) in zebrafish and *Table 1* summarizes the two-way ANOVA analysis. As expected, ACS decreased the distance total traveled, crossings, entries and time in the top area (*Figs. 4A–4D*, respectively) and increased the time in the bottom area (*Fig. 4E*). ALC (0.1, 1.0 and 10.0 mg/L) prevented the effects of ACS on the time in the top and bottom areas in the novel tank test (*Figs. 4D* and *4E*). Also, ALC (0.1 mg/L) prevented the effects of ACS on the total distance traveled.

Biochemical parameters

Figure 5 shows the effects of ALC (0.1, 1.0 and 10.0 mg/L) on oxidative status. ACS significantly increased lipid peroxidation (TBARS), non-protein sulfhydryl (NPSH) and superoxide dismutase (SOD) activity (*Figs. 5A, 5C* and *5D*, respectively), but did not alter sulfhydryl (SH) content and catalase (CAT) activity (*Figs. 5B* and *5E*, respectively). Treatment with ALC (0.1, 1.0 and 10.0 mg/L) prevented oxidative damage as measured by

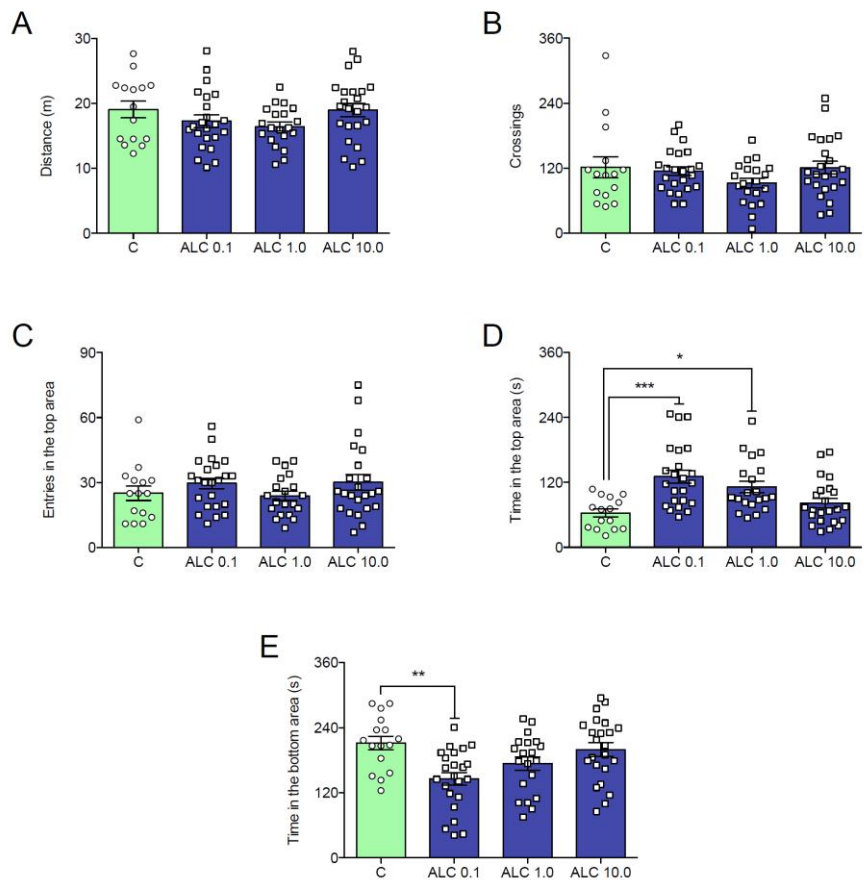


Figure 2 Effects of ALC (0.1, 1.0 and 10.0 mg/L) behavioral parameters in zebrafish submitted to the novel tank test. (A) distance traveled, (B) number of crossings, (C) entries and (D) time in the upper zone, and (E) time in the bottom zone. The data are presented as the mean + S.E.M. One-way ANOVA followed by Tukey post hoc test. $n = 15-23$. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. control group.

Full-size [DOI: 10.7717/peerj.5309/fig-2](https://doi.org/10.7717/peerj.5309/fig-2)

TBARS. ALC also prevented the increase of antioxidant defenses as measured by NPSH (0.1 mg/L) and SOD (0.1, 1.0 and 10.0 mg/L). Two-way ANOVA analyses were summarized in [Table 2](#).

DISCUSSION

Here, we showed for the first time that ALC presents anxiolytic effects in both novel tank and light/dark tests in zebrafish. Moreover, ALC was able to prevent the anxiogenic effects and lipid peroxidation induced by an acute stress protocol. These results indicate a potential use of ALC in mental disorders related to stress.

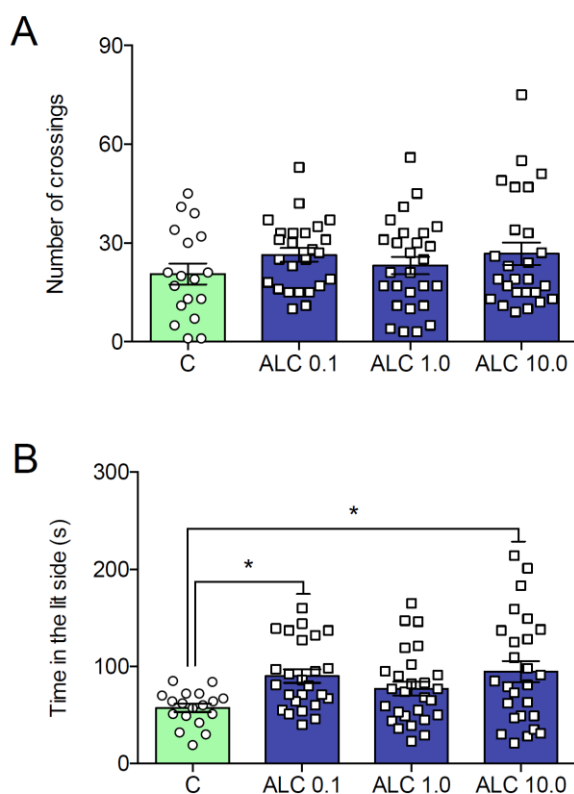


Figure 3 Effects of ALC (0.1, 1.0 and 10.0 mg/L) in the light/dark test in zebrafish. (A) number of crossings and (B) time in the lit side. The data are presented as the mean + S.E.M. One-way ANOVA followed by Tukey post hoc test. $n = 18-27$. * $p < 0.05$ vs. control group.

Full-size [DOI: 10.7717/peerj.5309/fig-3](https://doi.org/10.7717/peerj.5309/fig-3)

ALC increased the time spent in the upper as well as decreased the time spent in the bottom zones of the tank. Previous studies have shown that anxiolytic drugs such as buspirone, fluoxetine, diazepam, and ethanol increase the time spent in this zone (Bencan, Sledge & Levin, 2009; Egan et al., 2009; Gebauer et al., 2011). In the light/dark test, ALC increased the time spent in the lit side of the tank. This effect was observed with other drugs as clonazepam, bromazepam, diazepam, buspirone, and ethanol (Gebauer et al., 2011). Additionally, multi-target drugs other than ALC, for instance, N-acetylcysteine (NAC) and taurine, also increase the time in the lit side in the LTD in zebrafish (Mocelin et al., 2015; Mezzomo et al., 2015). In both NTT and LDT, ALC presented biphasic response. We can only speculate that different mechanisms of action may be involved in the effects of low versus high dose, but lower and higher concentrations would have to be tested for us to have a bigger picture of the dose–response relationship. ALC also prevented the locomotor impairment and anxiogenic behavior induced by the chasing stress protocol. Recently, our

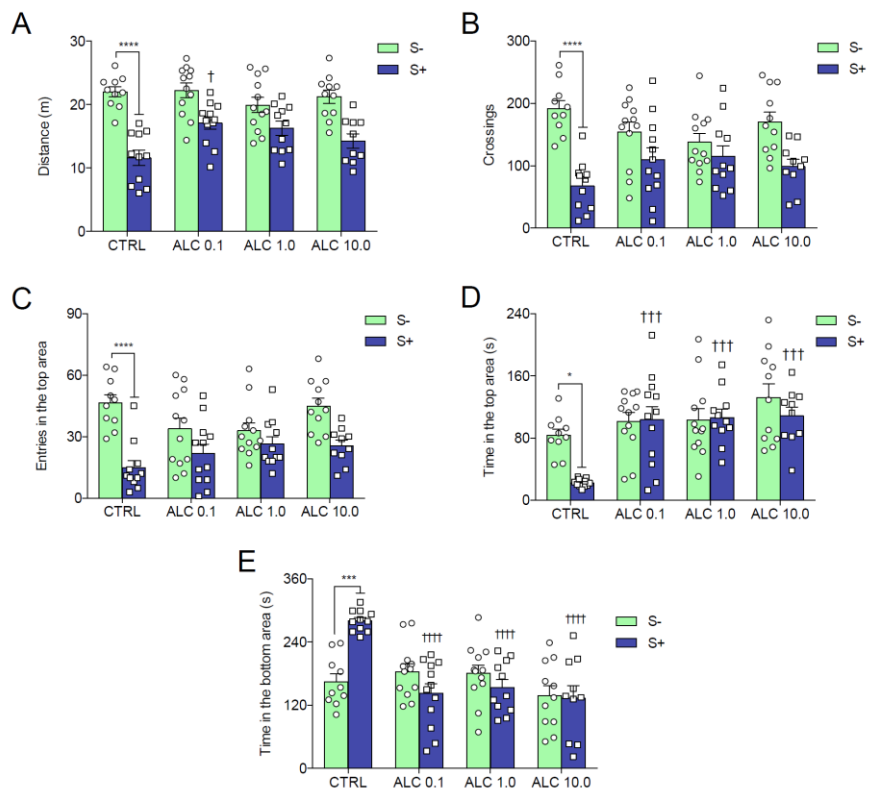


Figure 4 Effects of ALC pretreatment against stress-induced changes in behavioral parameters in zebrafish. (A) distance traveled, (B) number of crossings, (C) entries and (D) time in the upper zone, and (E) time in the bottom zone. The data are presented as the mean \pm S.E.M. Two-way ANOVA followed by Bonferroni's test. $n = 10-12$. * $p < 0.05$, *** $p < 0.001$, **** $p < 0.0001$ vs. control group (S-); † $p < 0.05$, †† $p < 0.001$, ††† $p < 0.0001$ vs. stressed control group (S+).

Full-size [DOI: 10.7717/peerj.5309/fig-4](https://doi.org/10.7717/peerj.5309/fig-4)

group has shown that fluoxetine, diazepam, and NAC prevented the effects of a similar stress protocol in zebrafish (Mocelin et al., 2015; Giacomini et al., 2016).

The anxiolytic and antidepressant effects of ALC have been already reported in rodents (Levine et al., 2005; Wang et al., 2015; Lau et al., 2017). ALC modulates the cholinergic system by increasing acetyl-CoA content and choline acetyltransferase activity. Moreover, it modulates GABAergic, dopaminergic and glutamatergic neurotransmitter systems (Chapela et al., 2009; Nasca et al., 2013; Wang et al., 2014; Singh et al., 2016; Chiechio, Canonico & Grilli, 2017). In rats, ALC decreased the immobility time in the forced swim test and increased sucrose preference in 3 days of treatment, whereas 14 days were necessary to obtain the same effects with clomipramine (Nasca et al., 2013).

Table 1 Results of two-way analysis of variance (ANOVA) of behavioral analysis and the interaction between treatment with ALC and acute chasing stress.

Dependent variable	Effects	F-value	DF	P-value
Total distance	Interaction	3.46	3,81	0.0201
	ALC	2.39	3,81	0.0745
	Stress	71.34	3,81	0.0001
Crossings	Interaction	4.04	3,81	0.0099
	ALC	0.10	3,81	0.9583
	Stress	37.11	3,81	0.0001
Entries in the top	Interaction	3.47	3,81	0.0198
	ALC	1.18	3,81	0.3215
	Stress	36.10	3,81	0.0001
Time in the top	Interaction	2.72	3,81	0.0499
	ALC	9.81	3,81	0.0001
	Stress	4.86	3,81	0.0303
Time in the bottom	Interaction	9.02	3,81	0.0001
	ALC	9.03	3,81	0.0001
	Stress	0.84	3,81	0.3613

Notes.

DF, degrees of freedom.

Significant effects ($p < 0.05$) are given in bold.

Under normal conditions, damage by reactive oxygen species (ROS) is kept in control by efficient antioxidant systems, such as SOD and CAT enzymes, as well as non-enzymatic scavengers (Schivone *et al.*, 2013; Schivone, Colaianna & Curtis, 2015; Sandi & Haller, 2015). Studies have demonstrated that ALC protects cells against lipid peroxidation and membrane breakdown through hydrogen peroxide scavenging (Kumaran *et al.*, 2003; Gülçin, 2006), and can promote the expression of antioxidant enzymes such as SOD and CAT (Augustyniak & Skrzydlewska, 2010; Li *et al.*, 2012).

Even though the ACS protocol increased antioxidant defenses (NPSH and SOD), it also caused lipid peroxidation (TBARS), which may indicate a possible adaptive response to ROS production during stressful conditions. Similar results were observed in zebrafish and reported in a previous study from our group using acute restraint stress (Dal Santo *et al.*, 2014). Even though detection of MDA levels by HPLC would be a more specific indicator of lipid peroxidation, the TBARS assay we used in this study has been reported by many previous articles using samples from zebrafish and other animals (Mihara & Uchiyama, 1978; Sunderman *et al.*, 1985; Armstrong & Browne, 1994; Yagi, 1998; Kim *et al.*, 2011; Basu *et al.*, 2014; Yavuzer *et al.*, 2016). The association of these factors could be related to the prevented effects of ALC, and that our results indicate a deficit in antioxidant defenses against lipid peroxidation in zebrafish submitted to the ACS protocol, providing

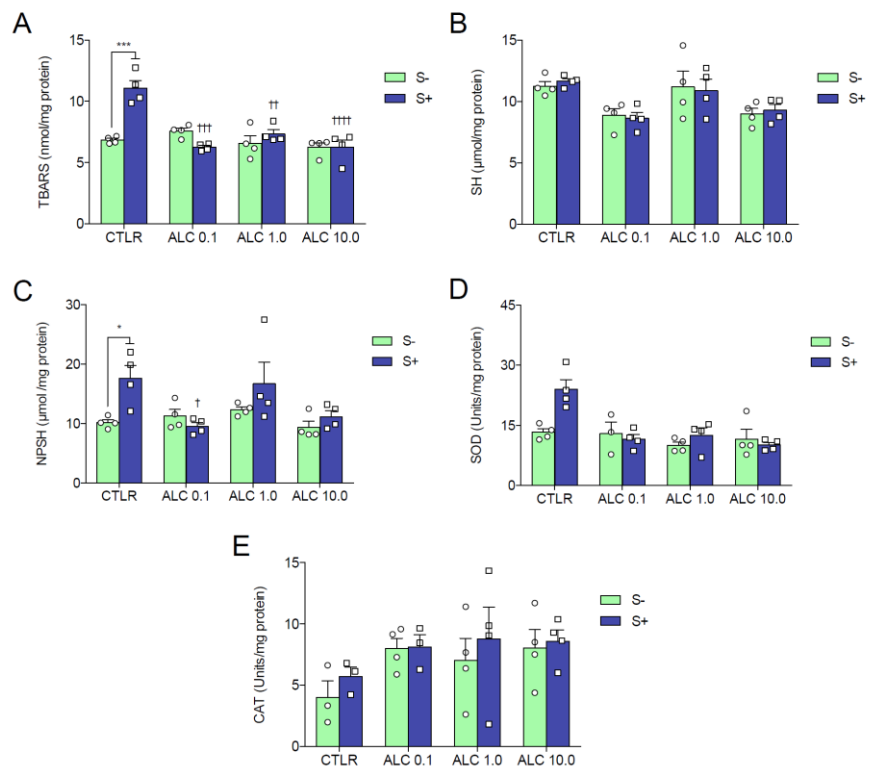


Figure 5 Effects of ALC pretreatment against stress-induced changes in biochemical parameters in zebrafish. (A) thiobarbituric acid reactive substances, (B) sulfhydryl, (C) non-protein sulphhydryl, (D) superoxide dismutase, and (E) catalase. The data are presented as the mean + S.E.M. Two-way ANOVA followed by Bonferroni's test. $n = 3-4$. * $p < 0.05$, ** $p < 0.001$ vs. control group (S-); † $p < 0.05$, †† $p < 0.001$, ††† $p < 0.0001$ vs. stressed control group (S+).

Full-size [DOI: 10.7717/peerj.5309/fig-5](https://doi.org/10.7717/peerj.5309/fig-5)

further evidence for the hypothesis of an association between behavior and ROS with the pathophysiology of mental disorders stress-related and their prevention by ALC.

CONCLUSION

ALC is already widely used as supplementation for people who want to lose weight/fat burner, but only a few studies assessed its effects on stress-related outcomes. In addition to its antioxidant actions, ALC is also able to restore mitochondrial function, which is relevant to combat the dysregulation of fatty acid metabolism in the mitochondria-associated with psychiatric disorders. Furthermore, there is evidence that ALC increases expression of metabotropic glutamate receptors via epigenetic mechanisms (Nasca *et al.*, 2013), which is also relevant for the pathophysiology of depression and other stress-related disorders.

Table 2 Results of two-way analysis of variance (ANOVA) of biochemical analysis and the interaction between treatment with ALC and acute chasing stress.

Dependent variable	Effects	F-value	DF	P-value
Lipid peroxidation (TBARS)	Interaction	14.70	3,24	0.0001
	ALC	14.39	3,24	0.0001
	Stress	8.80	1,24	0.0067
Sulphydryl (SH)	Interaction	0.14	3,24	0.9339
	ALC	7.80	3,24	0.0008
	Stress	0.01	1,24	0.9289
Non-protein thiol (NPSH)	Interaction	2.73	3,24	0.0665
	ALC	3.63	3,24	0.0273
	Stress	6.35	1,24	0.0188
Superoxide dismutase (SOD)	Interaction	5.46	3,23	0.0055
	ALC	9.93	3,23	0.0004
	Stress	4.26	1,23	0.0504
Catalase (CAT)	Interaction	0.13	3,21	0.9393
	ALC	1.89	3,21	0.1626
	Stress	0.87	1,21	0.3606

Notes.

DF, degrees of freedom.

Significant effects ($p < 0.05$) are given in bold font.

Our study adds to a growing body of literature demonstrating the role of antioxidants in modulating behavior and oxidative homeostasis. The data presented here thus warrants further investigation of ALC as a potential agent in the treatment of neuropsychiatric illness. Its novel mechanism of action and good tolerability also subsidize the additional studies necessary to assess its therapeutic potential in clinical settings.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico—Brazil (CNPq, No. 401162/2016-8 and 302800/2017-4). Riciery Mocelin and Matheus Marcon are recipients of a fellowship from Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Conselho Nacional de Desenvolvimento Científico e Tecnológico—Brazil: CNPq, No. 401162/2016-8, 302800/2017-4.

Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

Competing Interests

Angelo Piato is an Academic Editor for PeerJ.

Author Contributions

- Lais Pancotto performed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Ricieri Mocelin conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Matheus Marcon conceived and designed the experiments, performed the experiments, approved the final draft.
- Ana P. Herrmann conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, approved the final draft.
- Angelo Piato conceived and designed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

Animal Ethics

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

All experiments were approved by the Ethics Committee of Universidade Federal do Rio Grande do Sul (#30992/2015).

Data Availability

The following information was supplied regarding data availability:

The raw data is provided as [Data S1](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.5309#supplemental-information>.

REFERENCES

- Aebi H. 1984.** Catalase in vitro. *Methods in Enzymology* **105**:121–126.
- Armstrong D, Browne R. 1994.** The analysis of free radicals, lipid peroxides, antioxidant enzymes and compounds related to oxidative stress as applied to the clinical chemistry laboratory. *Advances in Experimental Medicine and Biology* **366**:43–58.
- Augustyniak A, Skrzydlewska E. 2010.** The influence of L-carnitine supplementation on the antioxidative abilities of serum and the central nervous system of ethanol-induced rats. *Metabolic Brain Disease* **25**:381–389 DOI [10.1007/s11011-010-9217-7](https://doi.org/10.1007/s11011-010-9217-7).
- Basu S, De D, Dev Khanna H, Kumar A. 2014.** Lipid peroxidation, DNA damage and total antioxidant status in neonatal hyperbilirubinemia. *Journal of Perinatology* **34**:519–523 DOI [10.1038/jp.2014.45](https://doi.org/10.1038/jp.2014.45).

- Bencan Z, Sledge D, Levin ED. 2009.** Buspirone, chlordiazepoxide and diazepam effects in a zebrafish model of anxiety. *Pharmacology, Biochemistry, and Behavior* **94**:75–80 DOI [10.1016/j.pbb.2009.07.009](https://doi.org/10.1016/j.pbb.2009.07.009).
- Berk M, Dean OM, Cotton SM, Jeavons S, Tanious M, Kohlmann K, Hewitt K, Moss K, Allwang C, Schapkaitz I, Robbins J, Cobb H, Ng F, Dodd S, Bush AI, Malhi GS. 2014.** The efficacy of adjunctive N-acetylcysteine in major depressive disorder: a double-blind, randomized, placebo-controlled trial. *The Journal of Clinical Psychiatry* **75**:628–636 DOI [10.4088/JCP.13m08454](https://doi.org/10.4088/JCP.13m08454).
- Chapela SP, Krieger N, Fernández EH, Stella CA. 2009.** Involvement of L-carnitine in cellular metabolism: beyond Acyl-CoA transport. *Mini Reviews in Medicinal Chemistry* **9**:1518–1526.
- Chiechio S, Canonico PL, Grilli M. 2017.** L-Acetylcarnitine: a mechanistically distinctive and potentially rapid-acting antidepressant drug. *International Journal of Molecular Sciences* **19**(1):1–13 DOI [10.3390/ijms19010011](https://doi.org/10.3390/ijms19010011).
- Cobb CA, Cole MP. 2015.** Oxidative and nitrate stress in neurodegeneration. *Neurobiology of Disease* **84**:4–21 DOI [10.1016/j.nbd.2015.04.020](https://doi.org/10.1016/j.nbd.2015.04.020).
- Dal Santo G, Conterato GMM, Barcellos LJG, Rosemberg DB, Piato AL. 2014.** Acute restraint stress induces an imbalance in the oxidative status of the zebrafish brain. *Neuroscience Letters* **558**:103–108 DOI [10.1016/j.neulet.2013.11.011](https://doi.org/10.1016/j.neulet.2013.11.011).
- Draper HH, Hadley M. 1990.** Malondialdehyde determination as index of lipid peroxidation. *Methods in Enzymology* **186**:421–431.
- Egan RJ, Bergner CL, Hart PC, Cachat JM, Canavello PR, Elegante MF, Elkhayat SI, Bartels BK, Tien AK, Tien DH, Mohnot S, Beeson E, Glasgow E, Amri H, Zukowska Z, Kalueff AV. 2009.** Understanding behavioral and physiological phenotypes of stress and anxiety in zebrafish. *Behavioural Brain Research* **205**:38–44 DOI [10.1016/j.bbr.2009.06.022](https://doi.org/10.1016/j.bbr.2009.06.022).
- Ellman GL. 1959.** Tissue sulfhydryl groups. *Archives of Biochemistry and Biophysics* **82**:70–77.
- Gebauer DL, Pagnussat N, Piato AL, Schaefer IC, Bonan CD, Lara DR. 2011.** Effects of anxiolytics in zebrafish: similarities and differences between benzodiazepines, buspirone and ethanol. *Pharmacology, Biochemistry, and Behavior* **99**:480–486 DOI [10.1016/j.pbb.2011.04.021](https://doi.org/10.1016/j.pbb.2011.04.021).
- Giacomini ACVV, Abreu MS, Giacomini LV, Siebel AM, Zimmerman FF, Rambo CL, Mocelin R, Bonan CD, Piato AL, Barcellos LJG. 2016.** Fluoxetine and diazepam acutely modulate stress induced-behavior. *Behavioural Brain Research* **296**:301–310 DOI [10.1016/j.bbr.2015.09.027](https://doi.org/10.1016/j.bbr.2015.09.027).
- Gülçin I. 2006.** Antioxidant and antiradical activities of L-carnitine. *Life Sciences* **78**:803–811 DOI [10.1016/j.lfs.2005.05.103](https://doi.org/10.1016/j.lfs.2005.05.103).
- Hassan W, Noreen H, Castro-Gomes V, Mohammadzai I, Da Rocha JBT, Landeira-Fernandez J. 2016.** Association of oxidative stress with psychiatric disorders. *Current Pharmaceutical Design* **22**:2960–2974.
- Khan KM, Collier AD, Meshalkina DA, Kysil EV, Khatsko SL, Kolesnikova T, Morzherin YY, Warnick JE, Kalueff AV, Echevarria DJ. 2017.** Zebrafish models in

- neuropsychopharmacology and CNS drug discovery. *British Journal of Pharmacology* **174**:1925–1944 DOI [10.1111/bph.13754](https://doi.org/10.1111/bph.13754).
- Kim H, Lee SW, Baek KM, Park JS, Min JH. 2011.** Continuous hypoxia attenuates paraquat-induced cytotoxicity in the human A549 lung carcinoma cell line. *Experimental & Molecular Medicine* **43**:494–500 DOI [10.3858/emm.2011.43.9.056](https://doi.org/10.3858/emm.2011.43.9.056).
- Kumaran S, Deepak B, Naveen B, Panneerselvam C. 2003.** Effects of levocarnitine on mitochondrial antioxidant systems and oxidative stress in aged rats. *Drugs in R&D* **4**:141–147.
- Lau T, Bigio B, Zelli D, McEwen BS, Nasca C. 2017.** Stress-induced structural plasticity of medial amygdala stellate neurons and rapid prevention by a candidate antidepressant. *Molecular Psychiatry* **22**:227–234 DOI [10.1038/mp.2016.68](https://doi.org/10.1038/mp.2016.68).
- Levin ED, Bencan Z, Cerutti DT. 2007.** Anxiolytic effects of nicotine in zebrafish. *Physiology & Behavior* **90**:54–58 DOI [10.1016/j.physbeh.2006.08.026](https://doi.org/10.1016/j.physbeh.2006.08.026).
- Levine J, Kaplan Z, Pettegrew JW, McClure RJ, Gershon S, Buriakovsky I, Cohen H. 2005.** Effect of intraperitoneal acetyl-L-carnitine (ALCAR) on anxiety-like behaviours in rats. *The International Journal of Neuropsychopharmacology* **8**:65–74 DOI [10.1017/S1461145704004596](https://doi.org/10.1017/S1461145704004596).
- Li J-L, Wang Q-Y, Luan H-Y, Kang Z-C, Wang C-B. 2012.** Effects of L-carnitine against oxidative stress in human hepatocytes: involvement of peroxisome proliferator-activated receptor alpha. *Journal of Biomedical Science* **19**:32 DOI [10.1186/1423-0127-19-32](https://doi.org/10.1186/1423-0127-19-32).
- Maes M, Galecki P, Chang YS, Berk M. 2011.** A review on the oxidative and nitrosative stress (O&NS) pathways in major depression and their possible contribution to the (neuro)degenerative processes in that illness. *Progress in Neuro-psychopharmacology & Biological Psychiatry* **35**:676–692 DOI [10.1016/j.pnpbp.2010.05.004](https://doi.org/10.1016/j.pnpbp.2010.05.004).
- Marcon M, Herrmann AP, Mocelin R, Rambo CL, Koakoski G, Abreu MS, Conterato GMM, Kist LW, Bogo MR, Zanatta L, Barcellos LJG, Piato AL. 2016.** Prevention of unpredictable chronic stress-related phenomena in zebrafish exposed to bromazepam, fluoxetine and nortriptyline. *Psychopharmacology* **233**(21–22):3815–3824 DOI [10.1007/s00213-016-4408-5](https://doi.org/10.1007/s00213-016-4408-5).
- Marcon M, Mocelin R, Benvenuto R, Costa T, Herrmann AP, De Oliveira DL, Koakoski G, Barcellos LJG, Piato A. 2018.** Environmental enrichment modulates the response to chronic stress in zebrafish. *The Journal of Experimental Biology* **22**:221 DOI [10.1242/jeb.176735](https://doi.org/10.1242/jeb.176735).
- Maximino C, Marques de Brito T, Dias CAG de M, Gouveia A, Morato S. 2010.** Scototaxis as anxiety-like behavior in fish. *Nature Protocols* **5**:209–216 DOI [10.1038/nprot.2009.225](https://doi.org/10.1038/nprot.2009.225).
- Mecocci P, Polidori MC. 2012.** Antioxidant clinical trials in mild cognitive impairment and Alzheimer's disease. *Biochimica et Biophysica Acta* **1822**:631–638 DOI [10.1016/j.bbadis.2011.10.006](https://doi.org/10.1016/j.bbadis.2011.10.006).

- Mezzomo NJ, Silveira A, Giuliani GS, Quadros VA, Rosemberg DB. 2015. The role of taurine on anxiety-like behaviors in zebrafish: a comparative study using the novel tank and the light-dark tasks. *Neuroscience Letters* **613**:19–24 DOI [10.1016/j.neulet.2015.12.037](https://doi.org/10.1016/j.neulet.2015.12.037).
- Mihara M, Uchiyama M. 1978. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. *Analytical Biochemistry* **86**:271–278.
- Misra HP, Fridovich I. 1972. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *The Journal of Biological Chemistry* **247**:3170–3175.
- Mocelin R, Herrmann AP, Marcon M, Rambo CL, Rohden A, Bevilaqua F, De Abreu MS, Zanatta L, Elisabetsky E, Barcellos LJG, Lara DR, Piato AL. 2015. N-acetylcysteine prevents stress-induced anxiety behavior in zebrafish. *Pharmacology, Biochemistry, and Behavior* **139**(Pt B):121–126 DOI [10.1016/j.pbb.2015.08.006](https://doi.org/10.1016/j.pbb.2015.08.006).
- Mocelin R, Marcon M, D’ambros S, Herrmann AP, Araujo AS da R, Piato A. 2017. Behavioral and biochemical effects of N-Acetylcysteine in zebrafish acutely exposed to ethanol. *Neurochemical Research* **43**(2):458–464 DOI [10.1007/s11064-017-2442-2](https://doi.org/10.1007/s11064-017-2442-2).
- Mocelin R, Marcon M, D’ambros S, Herrmann AP, Da Rosa Araujo AS, Piato A. 2018. Behavioral and biochemical effects of N-acetylcysteine in zebrafish acutely exposed to ethanol. *Neurochemical Research* **43**(2):458–464 DOI [10.1007/s11064-017-2442-2](https://doi.org/10.1007/s11064-017-2442-2).
- Nasca C, Bigio B, Zelli D, De Angelis P, Lau T, Okamoto M, Soya H, Ni J, Brichta L, Greengard P, Neve RL, Lee FS, McEwen BS. 2017. Role of the astroglial glutamate exchanger xct in ventral hippocampus in resilience to stress. *Neuron* **96**:402–413 DOI [10.1016/j.neuron.2017.09.020](https://doi.org/10.1016/j.neuron.2017.09.020).
- Nasca C, Xenos D, Barone Y, Caruso A, Scaccianoce S, Matrisciano F, Battaglia G, Mathé AA, Pittaluga A, Lionetto L, Simmaco M, Nicoletti F. 2013. L-acetylcarnitine causes rapid antidepressant effects through the epigenetic induction of mGlu2 receptors. *Proceedings of the National Academy of Sciences of the United States of America* **110**:4804–4809 DOI [10.1073/pnas.1216100110](https://doi.org/10.1073/pnas.1216100110).
- Ng F, Berk M, Dean O, Bush AI. 2008. Oxidative stress in psychiatric disorders: evidence base and therapeutic implications. *The International Journal of Neuropsychopharmacology* **11**:851–876 DOI [10.1017/S1461145707008401](https://doi.org/10.1017/S1461145707008401).
- Nicassio L, Fracasso F, Sirago G, Musicco C, Picca A, Marzetti E, Calvani R, Cantatore P, Gadaleta MN, Pesce V. 2017. Dietary supplementation with acetyl-L-carnitine counteracts age-related alterations of mitochondrial biogenesis, dynamics and antioxidant defenses in brain of old rats. *Experimental Gerontology* **98**:99–109 DOI [10.1016/j.exger.2017.08.017](https://doi.org/10.1016/j.exger.2017.08.017).
- Niedzielska E, Smaga I, Gawlik M, Moniczewski A, Stankowicz P, Pera J, Filip M. 2016. Oxidative stress in neurodegenerative diseases. *Molecular Neurobiology* **53**:4094–4125 DOI [10.1007/s12035-015-9337-5](https://doi.org/10.1007/s12035-015-9337-5).
- Ortiz GG, Pacheco Moisés FP, Mireles-Ramírez M, Flores-Alvarado LJ, González-Usigli H, Sánchez-González VJ, Sánchez-López AL, Sánchez-Romero L, Díaz-Barba EI, Santoscoy-Gutiérrez JF, Rivero-Moragrega P. 2017. Oxidative stress: love and

- hate history in central nervous system. *Advances in Protein Chemistry and Structural Biology* **108**:1–31 DOI [10.1016/bs.apcsb.2017.01.003](https://doi.org/10.1016/bs.apcsb.2017.01.003).
- Ozcan ME, Gulec M, Ozerol E, Polat R, Akyol O. 2004.** Antioxidant enzyme activities and oxidative stress in affective disorders. *International Clinical Psychopharmacology* **19**:89–95.
- Patel M. 2016.** Targeting oxidative stress in central nervous system disorders. *Trends in Pharmacological Sciences* **37**:768–778 DOI [10.1016/j.tips.2016.06.007](https://doi.org/10.1016/j.tips.2016.06.007).
- Pittman J, Piato A. 2017.** Developing zebrafish depression-related models. In: Kalueff AV, ed. *The rights and wrongs of zebrafish: behavioral phenotyping of zebrafish*. Cham: Springer International Publishing, 33–43 DOI [10.1007/978-3-319-33774-6_2](https://doi.org/10.1007/978-3-319-33774-6_2).
- Ribas GS, Vargas CR, Wajner M. 2014.** L-carnitine supplementation as a potential antioxidant therapy for inherited neurometabolic disorders. *Gene* **533**:469–476 DOI [10.1016/j.gene.2013.10.017](https://doi.org/10.1016/j.gene.2013.10.017).
- Sandi C, Haller J. 2015.** Stress and the social brain: behavioural effects and neurobiological mechanisms. *Nature Reviews Neuroscience* **16**:290–304 DOI [10.1038/nrn3918](https://doi.org/10.1038/nrn3918).
- Santos P, Herrmann AP, Benvenuti R, Noetzold G, Giongo F, Gama CS, Piato AL, Elisabetsky E. 2017.** Anxiolytic properties of N-acetylcysteine in mice. *Behavioural Brain Research* **317**:461–469 DOI [10.1016/j.bbr.2016.10.010](https://doi.org/10.1016/j.bbr.2016.10.010).
- Schiavone S, Colaianna M, Curtis L. 2015.** Impact of early life stress on the pathogenesis of mental disorders: relation to brain oxidative stress. *Current Pharmaceutical Design* **21**:1404–1412.
- Schiavone S, Jaquet V, Trabace L, Krause K-H. 2013.** Severe life stress and oxidative stress in the brain: from animal models to human pathology. *Antioxidants & Redox Signaling* **18**:1475–1490 DOI [10.1089/ars.2012.4720](https://doi.org/10.1089/ars.2012.4720).
- Sepand MR, Razavi-Azarkhiavi K, Omidi A, Zirak MR, Sabzevari S, Kazemi AR, Sabzevari O. 2016.** Effect of acetyl-L-carnitine on antioxidant status, lipid peroxidation, and oxidative damage of arsenic in rat. *Biological Trace Element Research* **171**:107–115 DOI [10.1007/s12011-015-0436-y](https://doi.org/10.1007/s12011-015-0436-y).
- Singh S, Mishra A, Mishra SK, Shukla S. 2017.** ALCAR promote adult hippocampal neurogenesis by regulating cell-survival and cell death-related signals in rat model of Parkinson's disease like-phenotypes. *Neurochemistry International* **108**:388–396 DOI [10.1016/j.neuint.2017.05.017](https://doi.org/10.1016/j.neuint.2017.05.017).
- Singh S, Mishra A, Srivastava N, Shukla R, Shukla S. 2016.** Acetyl-L-carnitine via upregulating dopamine D1 receptor and attenuating microglial activation prevents neuronal loss and improves memory functions in parkinsonian rats. *Molecular Neurobiology* **55**(1):583–602 DOI [10.1007/s12035-016-0293-5](https://doi.org/10.1007/s12035-016-0293-5).
- Smaga I, Niedzielska E, Gawlik M, Moniczewski A, Krzek J, Przegaliński E, Pera J, Filip M. 2015.** Oxidative stress as an etiological factor and a potential treatment target of psychiatric disorders. Part 2. Depression, anxiety, schizophrenia and autism. *Pharmacological Reports* **67**:569–580 DOI [10.1016/j.pharep.2014.12.015](https://doi.org/10.1016/j.pharep.2014.12.015).
- Stewart AM, Ullmann JFP, Norton WHJ, Parker MO, Brennan CH, Gerlai R, Kalueff AV. 2015.** Molecular psychiatry of zebrafish. *Molecular Psychiatry* **20**:2–17 DOI [10.1038/mp.2014.128](https://doi.org/10.1038/mp.2014.128).

- Sunderman FW, Marzouk A, Hopfer SM, Zaharia O, Reid MC. 1985.** Increased lipid peroxidation in tissues of nickel chloride-treated rats. *Annals of Clinical and Laboratory Science* **15**:229–236.
- Traina G. 2016.** The neurobiology of acetyl-L-carnitine. *Frontiers in Bioscience* **21**:1314–1329.
- Wang S-M, Han C, Lee S-J, Patkar AA, Masand PS, Pae C-U. 2014.** A review of current evidence for acetyl-L-carnitine in the treatment of depression. *Journal of Psychiatric Research* **53**:30–37 DOI [10.1016/j.jpsychires.2014.02.005](https://doi.org/10.1016/j.jpsychires.2014.02.005).
- Wang W, Lu Y, Xue Z, Li C, Wang C, Zhao X, Zhang J, Wei X, Chen X, Cui W, Wang Q, Zhou W. 2015.** Rapid-acting antidepressant-like effects of acetyl-L-carnitine mediated by PI3K/AKT/BDNF/VGF signaling pathway in mice. *Neuroscience* **285**:281–291 DOI [10.1016/j.neuroscience.2014.11.025](https://doi.org/10.1016/j.neuroscience.2014.11.025).
- Yagi K. 1998.** Simple assay for the level of total lipid peroxides in serum or plasma. *Methods in Molecular Biology* **108**:101–106 DOI [10.1385/0-89603-472-0:101](https://doi.org/10.1385/0-89603-472-0:101).
- Yavuzer H, Yavuzer S, Cengiz M, Erman H, Doventas A, Balci H, Erdinciler DS, Uzun H. 2016.** Biomarkers of lipid peroxidation related to hypertension in aging. *Hypertension Research* **39**:342–348 DOI [10.1038/hr.2015.156](https://doi.org/10.1038/hr.2015.156).

4 DISCUSSÃO E CONCLUSÃO

Nesse estudo mostramos pela primeira vez os efeitos ansiolíticos e antiestresse da acetil-L-carnitina (ALC) em peixes-zebra. A ALC aumentou o tempo de permanência na zona superior do aquário no teste de tanque novo e o tempo de permanência no lado claro no teste de claro/escuro. Nas concentrações utilizadas não foram observados efeitos sobre a atividade exploratória e locomoção. Além disso, animais submetidos ao estresse agudo por perseguição por rede apresentaram diminuição na distância percorrida, o número de cruzamentos e entradas e tempo de permanência na zona superior do aquário enquanto a ALC preveniu esses efeitos. Em relação aos parâmetros de status oxidativo, o estresse agudo induziu lipoperoxidação e aumentou os níveis de tióis não-proteicos e a atividade da enzima superóxido dismutase (SOD), sem afetar os níveis de sulfidrilas (SH) e da catalase (CAT). O tratamento com ALC preveniu a lipoperoxidação e o aumento da superóxido dismutase.

No teste de tanque novo, o tempo de permanência e o número de entradas na zona superior do aquário é frequentemente utilizado como uma medida de ansiedade. Ou seja, intervenções ansiogênicas como o estresse agudo por perseguição (de Abreu et al., 2014; Giacomini et al., 2016) ou contenção (Piato et al., 2011) diminuem tanto o tempo como o número de entradas na zona superior, enquanto ansiolíticos como a fluoxetina, a buspirona, o diazepam e o etanol aumentam o tempo de permanência na zona superior e o número de entradas por reduzir a ansiedade (Bencan et al., 2009; Gebauer et al., 2011; Maximino et al., 2014).

No teste de claro/escuro, a ALC aumentou o tempo de permanência no lado claro do aparato. Esse efeito foi observado com outros ansiolíticos como clonazepam, bromazepam, diazepam, buspirona e etanol (Gebauer et al., 2011; Maximino et al.,

2014). Do mesmo modo, compostos com ação multialvo como a taurina e N-acetilcisteína também aumentam o tempo de permanência no lado claro do aparato (Mezzomo et al., 2016; Mocelin et al., 2015).

Em condições normais, o dano por espécies reativas de oxigênio (EROs) é prevenido por eficientes sistemas antioxidantes enzimáticos como SOD e a CAT e não-enzimáticos (Sandi and Haller, 2015; Schiavone et al., 2012). Estudos demonstraram que o ALC protege as células contra a peroxidação lipídica e a degradação da membrana por meio da depuração de peróxido de hidrogênio (Gülçin, 2006) e aumentou a expressão de enzimas antioxidantes como a SOD e a CAT (Augustyniak and Skrzydlewska, 2010). Embora o protocolo de estresse agudo tenha aumentado as defesas antioxidantes (NPSH e SOD), também causou a peroxidação lipídica (TBARS), o que pode indicar uma possível resposta adaptativa à produção de ROS durante condições estressantes. Resultados semelhantes foram observados em peixe-zebra e relatados em um estudo anterior do nosso grupo usando estresse por contenção (Dal Santo et al., 2014).

Os dados obtidos nessa dissertação corroboram com os disponíveis na literatura e agregam informações sobre os efeitos desse composto em diferentes modelos de ansiedade em peixes-zebra bem como sobre o potencial mecanismo de neuromodulação da ALC. Entretanto, mais estudos são necessários para ampliar a caracterização dos efeitos da ALC sobre transtornos de ansiedade bem como melhorar a caracterização do mecanismo de ação.

REFERÊNCIAS

- Amaral, A.D., 2014. Comparação entre SNRI e SSRI na indução da remissão da perturbação depressiva major: uma revisão baseada na evidência. *Rev. Port. Med. Geral E Fam.* 30, 174–180.
- Andreatini, R., Boerngen-Lacerda, R., Zorzetto Filho, D., 2001. Tratamento farmacológico do transtorno de ansiedade generalizada: perspectivas futuras. *Rev. Bras. Psiquiatr.* 23, 233–242. <https://doi.org/10.1590/S1516-44462001000400011>
- Augustyniak, A., Skrzydlewska, E., 2010. The influence of L-carnitine supplementation on the antioxidative abilities of serum and the central nervous system of ethanol-induced rats. *Metab. Brain Dis.* 25, 381–389. <https://doi.org/10.1007/s11011-010-9217-7>
- Bencan, Z., Sledge, D., Levin, E.D., 2009. Buspirone, chlordiazepoxide and diazepam effects in a zebrafish model of anxiety. *Pharmacol. Biochem. Behav.* 94, 75–80. <https://doi.org/10.1016/j.pbb.2009.07.009>
- Braga, J.E.F., Pordeus, L.C., Silva, A.T.M.C., Pimenta, F.C.F., Diniz, M. de F.F.M., Almeida, R.N., 2010. Ansiedade Patológica: Bases Neurais e Avanços na Abordagem Psicofarmacológica. *Rev Bras Ciênc Saúde.*
- Brass, E.P., 2000. Supplemental carnitine and exercise. *Am. J. Clin. Nutr.* 72, 618S–23S.
- Calvani, M., Arrigoni-Martelli, E., 1999. Attenuation by acetyl-L-carnitine of neurological damage and biochemical derangement following brain ischemia and reperfusion. *Int. J. Tissue React.* 21, 1–6.

- Castillo, A.R.G., Recondo, R., Asbahr, F.R., Manfro, G.G., 2000. Transtornos de ansiedad. *Rev. Bras. Psiquiatr.* 22, 20–23. <https://doi.org/10.1590/S1516-44462000000600006>
- Chapela, S.P., Kriguer, N., Fernández, E.H., Stella, C.A., 2009. Involvement of L-carnitine in cellular metabolism: beyond Acyl-CoA transport. *Mini Rev. Med. Chem.* 9, 1518–1526.
- Craske, M.G., Rauch, S.L., Ursano, R., Prenoveau, J., Pine, D.S., Zinbarg, R.E., 2009. What is an anxiety disorder? *Depress. Anxiety* 26, 1066–1085. <https://doi.org/10.1002/da.20633>
- Cuccurazzu, B., Bortolotto, V., Valente, M.M., Ubezio, F., Koverech, A., Canonico, P.L., Grilli, M., 2013. Upregulation of mGlu2 receptors via NF- κ B p65 acetylation is involved in the Proneurogenic and antidepressant effects of acetyl-L-carnitine. *Neuropsychopharmacol. Off. Publ. Am. Coll. Neuropsychopharmacol.* 38, 2220–2230. <https://doi.org/10.1038/npp.2013.121>
- de Abreu, M.S., Koakoski, G., Ferreira, D., Oliveira, T.A., da Rosa, J.G.S., Gusso, D., Giacomini, A.C.V., Piato, A.L., Barcellos, L.J.G., 2014. Diazepam and fluoxetine decrease the stress response in zebrafish. *PLoS One* 9, e103232.
- Di Cesare Mannelli, L., Ghelardini, C., Toscano, A., Pacini, A., Bartolini, A., 2010. The neuropathy-protective agent acetyl-L-carnitine activates protein kinase C-gamma and MAPKs in a rat model of neuropathic pain. *Neuroscience* 165, 1345–1352. <https://doi.org/10.1016/j.neuroscience.2009.11.021>
- Drake, J., Kanski, J., Varadarajan, S., Tsoras, M., Butterfield, D.A., 2002. Elevation of brain glutathione by gamma-glutamylcysteine ethyl ester protects against peroxynitrite-induced oxidative stress. *J. Neurosci. Res.* 68, 776–784. <https://doi.org/10.1002/jnr.10266>

- Fariello, R.G., Ferraro, T.N., Golden, G.T., DeMattei, M., 1988. Systemic acetyl-L-carnitine elevates nigral levels of glutathione and GABA. *Life Sci.* 43, 289–292.
- Galvão-Coelho, N.L., Silva, H.P.A., Sousa, M.B.C. de, Galvão-Coelho, N.L., Silva, H.P.A., Sousa, M.B.C. de, 2015. Stress response: II. Resilience and vulnerability. *Estud. Psicol. Natal* 20, 72–81. <https://doi.org/10.5935/1678-4669.20150009>
- Gebauer, D.L., Pagnussat, N., Piato, Â.L., Schaefer, I.C., Bonan, C.D., Lara, D.R., 2011. Effects of anxiolytics in zebrafish: Similarities and differences between benzodiazepines, buspirone and ethanol. *Pharmacol. Biochem. Behav.* 99, 480–486. <https://doi.org/10.1016/j.pbb.2011.04.021>
- Giacomini, A.C.V.V., Abreu, M.S., Giacomini, L.V., Siebel, A.M., Zimmerman, F.F., Rambo, C.L., Mocelin, R., Bonan, C.D., Piato, A.L., Barcellos, L.J.G., 2016. Fluoxetine and diazepam acutely modulate stress induced-behavior. *Behav. Brain Res.* 296, 301–310. <https://doi.org/10.1016/j.bbr.2015.09.027>
- Griebel, G., Holmes, A., 2013. 50 years of hurdles and hope in anxiolytic drug discovery. *Nat. Rev. Drug Discov.* 12, 667–687. <https://doi.org/10.1038/nrd4075>
- Gülçin, İ., 2006. Antioxidant and antiradical activities of L-carnitine. *Life Sci.* 78, 803–811. <https://doi.org/10.1016/j.lfs.2005.05.103>
- Holmes, A., Heilig, M., Rupniak, N.M.J., Steckler, T., Griebel, G., 2003. Neuropeptide systems as novel therapeutic targets for depression and anxiety disorders. *Trends Pharmacol. Sci.* 24, 580–588. <https://doi.org/10.1016/j.tips.2003.09.011>

- Jones, L.L., McDonald, D.A., Borum, P.R., 2010. Acylcarnitines: role in brain. *Prog. Lipid Res.* 49, 61–75. <https://doi.org/10.1016/j.plipres.2009.08.004>
- Kido, Y., Tamai, I., Ohnari, A., Sai, Y., Kagami, T., Nezu, J., Nikaido, H., Hashimoto, N., Asano, M., Tsuji, A., 2001. Functional relevance of carnitine transporter OCTN2 to brain distribution of L-carnitine and acetyl-L-carnitine across the blood-brain barrier. *J. Neurochem.* 79, 959–969.
- Lau, T., Bigio, B., Zelli, D., McEwen, B.S., Nasca, C., 2016. Stress-induced structural plasticity of medial amygdala stellate neurons and rapid prevention by a candidate antidepressant. *Mol. Psychiatry.* <https://doi.org/10.1038/mp.2016.68>
- Leombruni, P., Miniotti, M., Colonna, F., Sica, C., Castelli, L., Bruzzone, M., Parisi, S., Fusaro, E., Sarzi-Puttini, P., Atzeni, F., Torta, R.G., 2015. A randomised controlled trial comparing duloxetine and acetyl L-carnitine in fibromyalgic patients: preliminary data. *Clin. Exp. Rheumatol.* 33, S82-85.
- Levine, J., Kaplan, Z., Pettegrew, J.W., McClure, R.J., Gershon, S., Buriakovsky, I., Cohen, H., 2005. Effect of intraperitoneal acetyl-L-carnitine (ALCAR) on anxiety-like behaviours in rats. *Int. J. Neuropsychopharmacol.* 8, 65–74. <https://doi.org/10.1017/S1461145704004596>
- Levitan, M.N., Chagas, M.H., Linares, I.M., Crippa, J.A., Terra, M.B., Giglio, A.T., Cordeiro, J.L.C., Garcia, G.J., Hasan, R., Andrada, N.C., Nardi, A.E., Levitan, M.N., Chagas, M.H., Linares, I.M., Crippa, J.A., Terra, M.B., Giglio, A.T., Cordeiro, J.L.C., Garcia, G.J., Hasan, R., Andrada, N.C., Nardi, A.E., 2013. Brazilian Medical Association guidelines for the diagnosis and differential diagnosis of panic disorder. *Rev. Bras. Psiquiatr.* 35, 406–415. <https://doi.org/10.1590/1516-4446-2012-0860>

Maes, M., Galecki, P., Chang, Y.S., Berk, M., 2011. A review on the oxidative and nitrosative stress (O&NS) pathways in major depression and their possible contribution to the (neuro)degenerative processes in that illness. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 35, 676–692.
<https://doi.org/10.1016/j.pnpbp.2010.05.004>

Marcon, M., Herrmann, A.P., Mocelin, R., Rambo, C.L., Koakoski, G., Abreu, M.S., Conterato, G.M.M., Kist, L.W., Bogo, M.R., Zanatta, L., Barcellos, L.J.G., Piato, A.L., 2016. Prevention of unpredictable chronic stress-related phenomena in zebrafish exposed to bromazepam, fluoxetine and nortriptyline. *Psychopharmacology (Berl.)* 233, 3815–3824. <https://doi.org/10.1007/s00213-016-4408-5>

Marcon, M., Mocelin, R., de Oliveira, D.L., da Rosa Araujo, A.S., Herrmann, A.P., Piato, A., 2019. Acetyl-L-carnitine as a putative candidate for the treatment of stress-related psychiatric disorders: Novel evidence from a zebrafish model. *Neuropharmacology* 150, 145–152.
<https://doi.org/10.1016/j.neuropharm.2019.03.024>

Maximino, C., Silva, A.W.B. da, Araújo, J., Lima, M.G., Miranda, V., Puty, B., Benzecry, R., Picanço-Diniz, D.L.W., Jr, A.G., Oliveira, K.R.M., Herculano, A.M., 2014. Fingerprinting of Psychoactive Drugs in Zebrafish Anxiety-Like Behaviors. *PLOS ONE* 9, e103943.
<https://doi.org/10.1371/journal.pone.0103943>

McEwen, B.S., Morrison, J.H., 2013. The Brain on Stress: Vulnerability and Plasticity of the Prefrontal Cortex over the Life Course. *Neuron* 79, 16–29.
<https://doi.org/10.1016/j.neuron.2013.06.028>

- Mezzomo, N.J., Silveira, A., Giuliani, G.S., Quadros, V.A., Rosemberg, D.B., 2016. The role of taurine on anxiety-like behaviors in zebrafish: A comparative study using the novel tank and the light–dark tasks. *Neurosci. Lett.* 613, 19–24.
- Mocelin, R., Herrmann, A.P., Marcon, M., Rambo, C.L., Rohden, A., Bevilaqua, F., de Abreu, M.S., Zanatta, L., Elisabetsky, E., Barcellos, L.J.G., Lara, D.R., Piato, A.L., 2015. N-acetylcysteine prevents stress-induced anxiety behavior in zebrafish. *Pharmacol. Biochem. Behav.*
<https://doi.org/10.1016/j.pbb.2015.08.006>
- Nasca, C., Bigio, B., Lee, F.S., Young, S.P., Kautz, M.M., Albright, A., Beasley, J., Millington, D.S., Mathé, A.A., Kocsis, J.H., Murrough, J.W., McEwen, B.S., Rasgon, N., 2018. Acetyl-L-carnitine deficiency in patients with major depressive disorder. *Proc. Natl. Acad. Sci.* 115, 8627–8632.
<https://doi.org/10.1073/pnas.1801609115>
- Nasca, C., Xenos, D., Barone, Y., Caruso, A., Scaccianoce, S., Matrisciano, F., Battaglia, G., Mathé, A.A., Pittaluga, A., Lionetto, L., Simmaco, M., Nicoletti, F., 2013. L-acetylcarnitine causes rapid antidepressant effects through the epigenetic induction of mGlu2 receptors. *Proc. Natl. Acad. Sci. U. S. A.* 110, 4804–4809. <https://doi.org/10.1073/pnas.1216100110>
- Ng, F., Berk, M., Dean, O., Bush, A.I., 2008. Oxidative stress in psychiatric disorders: evidence base and therapeutic implications. *Int. J. Neuropsychopharmacol.* 11, 851–876. <https://doi.org/10.1017/S1461145707008401>
- Nordon, D.G., Akamine, K., Novo, N.F., Hübner, C. von K., 2009. Characteristics of the use of benzodiazepines by women seeking treatment in primary care. *Rev. Psiquiatr. Rio Gd. Sul* 31, 152–158. <https://doi.org/10.1590/S0101-81082009000300004>

- Pettegrew, J.W., Levine, J., Gershon, S., Stanley, J.A., Servan-Schreiber, D., Panchalingam, K., McClure, R.J., 2002. 31P-MRS study of acetyl-L-carnitine treatment in geriatric depression: preliminary results. *Bipolar Disord.* 4, 61–66.
- Piato, A.L., Rosemberg, D.B., Capiotti, K.M., Siebel, A.M., Herrmann, A.P., Ghisleni, G., Vianna, M.R., Bogo, M.R., Lara, D.R., Bonan, C.D., 2011. Acute restraint stress in zebrafish: behavioral parameters and purinergic signaling. *Neurochem. Res.* 36, 1876.
- Portugal, F.B., Campos, M.R., Gonçalves, D.A., Mari, J. de J., Fortes, S.L.C.L., Portugal, F.B., Campos, M.R., Gonçalves, D.A., Mari, J. de J., Fortes, S.L.C.L., 2016. Quality of life of primary care patients in Rio de Janeiro and São Paulo, Brasil: associations with stressful life events and mental health. *Ciênc. Amp Saúde Coletiva* 21, 497–508. <https://doi.org/10.1590/1413-81232015212.20032015>
- Robinson, B.L., Dumas, M., Ali, S.F., Paule, M.G., Gu, Q., Kanungo, J., 2018. Mechanistic studies on ketamine-induced mitochondrial toxicity in zebrafish embryos. *Neurotoxicol. Teratol.* 69, 63–72. <https://doi.org/10.1016/j.ntt.2017.12.005>
- Sandi, C., Haller, J., 2015. Stress and the social brain: behavioural effects and neurobiological mechanisms. *Nat. Rev. Neurosci.* 16, 290. <https://doi.org/10.1038/nrn3918>
- Schalleberger, J.B., Colet, C. de F., Schalleberger, J.B., Colet, C. de F., 2016. Assessment of dependence and anxiety among benzodiazepine users in a provincial municipality in Rio Grande do Sul, Brazil. *Trends Psychiatry Psychother.* 38, 63–70. <https://doi.org/10.1590/2237-6089-2015-0041>

- Schiavone, S., Jaquet, V., Trabace, L., Krause, K.-H., 2012. Severe Life Stress and Oxidative Stress in the Brain: From Animal Models to Human Pathology. *Antioxid. Redox Signal.* 18, 1475–1490. <https://doi.org/10.1089/ars.2012.4720>
- Silva, M.G.F., Fernandes, C.P., Santos, T.C. da S., Silva, T.L.P. da, 2012. Oral supplementation of L-carnitine combined with exercise and respiratory training in patients with chronic obstructive pulmonary disease: preliminary study. *Fisioter. E Pesqui.* 19, 320–325. <https://doi.org/10.1590/S1809-29502012000400005>
- Smeland, O.B., Meisingset, T.W., Borges, K., Sonnewald, U., 2012. Chronic acetyl-L-carnitine alters brain energy metabolism and increases noradrenaline and serotonin content in healthy mice. *Neurochem. Int.* 61, 100–107. <https://doi.org/10.1016/j.neuint.2012.04.008>
- Steiber, A., Kerner, J., Hoppel, C.L., 2004. Carnitine: a nutritional, biosynthetic, and functional perspective. *Mol. Aspects Med.* 25, 455–473. <https://doi.org/10.1016/j.mam.2004.06.006>
- Wang, S.-M., Han, C., Lee, S.-J., Patkar, A.A., Masand, P.S., Pae, C.-U., 2014. A review of current evidence for acetyl-L-carnitine in the treatment of depression. *J. Psychiatr. Res.* 53, 30–37. <https://doi.org/10.1016/j.jpsychires.2014.02.005>